IndiMag Pathogen Kit
Quick Start Protocol

For use with magnetic particle processors (e.g. KingFisher Flex)

The IndiMag Pathogen Kit (cat. no. SP947457) and the IndiMag Pathogen Kit w/o plastics (cat. no. SP947257) can be stored at room temperature (15-25°C).
For expiry date information, read the label on the kit box.

Further information and support

- IndiMag Pathogen Kit or IndiMag Pathogen Kit w/o plastics Handbook: **www.indical.com/handbooks**
- Technical assistance: **support@indical.com**

Important notes before starting

- Read the safety information in the instrument user manual before use.
- Dissolve carrier RNA in Buffer AVE as indicated on the tube.
- Add isopropanol (100%) to Buffer ACB and ethanol (96-100%) to Buffers AW1 and AW2 before use. See the respective bottle labels for volumes.
- Vortex MagAttract Suspension G for 3 minutes and ensure that it is fully resuspended.
- Equilibrate buffers to room temperature (15-25ºC).
- If using a pack of 2 magnetic head covers, store the second cover in another 96-well deep well plate. Take care to not bend the covers.

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Procedure

1. Label and prepare 4 x 96-well deep well plates and 1 x 96-well microplate (slots 2-6) according to Table 1.

Table 1: Instrument setup and reagent volumes

Slot	Loading message	Format	Item to add	Volume per well
6	Load Rod Cover	96-well deep well plate	Cover for 96 tip comb	—
5	Load Elution	96-well microplate	Buffer AVE	100 µl
4	Load Wash 3	96-well deep well plate	Ethanol (96–100%)	750 µl
3	Load Wash 2	96-well deep well plate	Buffer AW2	700 µl
2	Load Wash 1	96-well deep well plate	Buffer AW1	700 µl
1	Load Lysate	96-well deep well plate	Lysate*	720 µl

* Includes 20 µl Proteinase K, 200 µl sample and 500 µl Buffer VXL mixture

2. Prepare Buffer VXL mixture according to Table 2.

Table 2: Buffer VXL mixture preparation

Reagent	1 reaction	48 reactions	96 reactions
Buffer VXL	100 µl	4.8 ml	9.6 ml
Buffer ACB	400 µl	19.2 ml	38.4 ml
MagAttract Suspension G	25 µl	1.2 ml	2.4 ml
Carrier RNA (1 µg/µl)	1 µl	48 µl	96 µl

* Prepared volumes are 105% of required volumes to compensate for pipetting errors and possible evaporation. Excess buffer should be discarded.

3. Pipet 20 µl Proteinase K into the bottom of a new 96-well deep well plate well and add 200 µl sample (slot 1) according to Table 1.

   Note: If your sample volume is less than 200 µl, bring it to 200 µl by adding PBS.

4. Mix Buffer VXL mixture thoroughly for 30 s and add 500 µl Buffer VXL mixture to each sample in the 96-well deep well plate.

5. Immediately load prepared plates onto the processor and start appropriate script.